* Could you share a little more information on the specifics of the application, i.e. what the heater is going into and what you are trying to achieve?
* Do you have a sketch of the heater you can share?
* Have you already determined the input voltage and required wattage?
* How do you plan on adhering the heater to your part?
* How many prototypes will you need and how quickly would you need them?
* What is your estimated annual usage in production?

12-15-22 AutoCyPlex Full Auto Test

Cycles (3x)

1. Cycle 1
   1. Villin-488
   2. PDGF-555
   3. Gamma-actin-647
2. Cycle 2
   1. Ezrin—488
   2. PDGF-555
   3. Na/K ATPase -647
3. Cycle 3
   1. PCNA-488
   2. RAB7-594
   3. GLUT2-647

Notes

* Tiling is snake style that starts in upper left corner and goes right, down, left down, etc. and is always rectilinear
* All antibodies are used in 1:100 [C] and are prepared with 350uL
* mCPBA is used in [C] of 5mM for 5 minutes. Prepared from 1M stock in ethanol made fresh on 12-15-22
* All antibodies diluted in blocking buffer that is odyssey blocking with 0.1% triton x-100

Steps to Take

1. Mount to slide and install pressure plate.
2. Mount to tilt stage
3. Tighten coupler set screws
4. Flow fluid until certain it is good and flowing well and not leaking
5. Find tissue
6. Adjust to kinda kohler excitation
7. Tweak tilt to get pretty flat
8. Define MM surface
9. Run

Notes

* New chamber that doesn’t leak due to not drilling holes, but printing them.
* Cy1 = real cycle 2 dyes
* Cy2 = bleached after cy1
* Cy3 = real cy3 dyes
* Cy4 = bleached after cy3
* Cy5 = real cy1 dyes
* Big bubble stuck after H2O2 based bleaching.